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**{Exhibit 12}**

**Boguslaski et al., U.S. Patent 4,230,797, issued  
October 28, 1980**



**United States Patent** [11]**4,230,797****Boguslaski et al.**

[45]

**Oct. 28, 1980****[54] HETEROGENOUS SPECIFIC BINDING ASSAY EMPLOYING A COENZYME AS LABEL****[75] Inventors:** Robert C. Boguslaski, Elkhart; Robert J. Carrico, Bremen, both of Ind.; James E. Christner, Ann Arbor, Mich.**[73] Assignee:** Miles Laboratories, Inc., Elkhart, Ind.**[21] Appl. No.:** 894,838**[22] Filed:** Apr. 10, 1978**Related U.S. Application Data****[63]** Continuation of Ser. No. 667,982, Mar. 18, 1976, abandoned, which is a continuation-in-part of Ser. No. 572,008, Apr. 28, 1975, abandoned.**[51] Int. Cl.<sup>2</sup> .....** C12Q 1/66**[52] U.S. Cl. ....** 435/7; 23/230 B; 424/12; 435/810**[58] Field of Search .....** 23/230 B; 195/99, 103.5 A, 195/103.5 R; 424/8, 12; 435/7, 810**[56] References Cited****U.S. PATENT DOCUMENTS**

3,659,104	4/1972	Gross et al. ....	23/230 B
3,817,837	6/1974	Rubenstein et al. ....	195/103.5 R
3,839,153	10/1974	Schuurs et al. ....	424/12
3,880,934	4/1975	Rammler ....	195/103.5 A
3,998,943	12/1976	Ullman et al. ....	424/12

**OTHER PUBLICATIONS**

Miles, et al., "Labelled Antibodies and Immunological Assay Systems", *Nature*, vol. 219 (1968), pp. 186-189.

Sercarz, et al., "Antigen Binding to Cells: Determination by Enzymic Fluorogenic Group Hydrolysis", *Science*, vol. 159 (1968), pp. 884-885.

Zinchuk, et al., "Chemiluminescent of Determination of

Microamounts of Hydrogen Peroxide", *Chem. Abst.*, vol. 79, No. 2 (1973), p 571, Abs. No. 13219.

Slawinski, et al., "Use of Chemiluminescence for Determination of Peroxidase Activity in Vegetable Extracts III, Determination of Peroxidase Content in Vegetables and Fruits", *Chem. Absts.*, vol. 80, No. 3 (1974), p. 144, Abs. No. 11635p.

*Primary Examiner*—Thomas G. Wiseman*Attorney, Agent, or Firm*—Andrew L. Klawitter**[57]****ABSTRACT**

An improved heterogeneous specific binding assay method which employs a substance having reactant activity, i.e., a reactant, as a labeling substance in the detection of a ligand in a liquid medium. The method is carried out using reagent means which comprises, as its labeled constituent, a conjugate formed of a specific binding substance coupled to the reactant. The reactant advantageously is an enzymatic reactant such as an enzyme substrate or coenzyme. The activity of the conjugated reactant as a constituent of a predetermined reaction system is utilized as means for monitoring the extent of binding of the labeled constituent in conventional heterogeneous specific binding assay schemes. The presence of a ligand in a liquid medium may be determined following conventional competitive binding manipulative techniques. After the necessary separation of the bound and free phases resulting in the specific binding reaction system, the extent of binding of the labeled constituent is determined by contacting either phase with the necessary materials to form the predetermined monitoring reaction system in which the labeling substance is active and assessing reactant activity therein.

**26 Claims, No Drawings**



## HETEROGENOUS SPECIFIC BINDING ASSAY EMPLOYING A COENZYME AS LABEL

### CROSS-REFERENCE TO RELATED APPLICATION

This is a continuation of application Ser. No. 667,982, filed Mar. 18, 1976, now abandoned, which was a continuation-in-part of application Ser. No. 572,008, filed Apr. 28, 1975, now abandoned.

### BACKGROUND OF THE INVENTION

#### 1. Field of the Invention

This invention relates to methods and means for determining the presence of a ligand in a liquid medium based on the affinity of the ligand for a specific binding partner thereof. In particular, this invention relates to methods and means for use in specific binding assays which do not employ radioactive materials or modified enzymes as the labeling substance.

The desirability of a convenient, reliable, and nonhazardous means for detecting the presence of low concentrations of substances in liquids is self-evident. This is particularly true in the field of clinical chemistry where constituents of body fluids which may appear in concentrations as low as  $10^{-11}$  molar are known to be of pathological significance. The difficulty of detecting such low concentrations is compounded in the field of clinical chemistry where sample size is usually quite limited.

Classically, substances have been detected in liquids based on a reaction scheme wherein the substance to be detected is a necessary reactant. The presence of unknown is indicated by the appearance of a reaction product or the disappearance of a known reactant. In certain instances, such an assay method may be quantitative, based on a measurement of either the rate of appearance of product or disappearance of reactant or measurement of the aggregate amount of product produced or reactant consumed in attaining equilibrium. Each assay reaction system is necessarily either limited to use in the detection of only a small group of substances or is non-specific.

#### 2. Description of the Prior Art

The search for assay systems which are highly specific yet adaptable to the detection of a wide range of substances has evolved the radioimmunoassay. In this system a known amount of a radiolabeled form of the substance to be detected is allowed to compete with the unknown for a limited quantity of antibody specific for the unknown. The amount of the labeled form that becomes bound to antibody varies inversely with the level of unknown present. Inherent in the radioimmunoassay technique is the need to separate the labeled form of substance to be detected which becomes bound to antibody, the bound phase, from that which does not become so bound, the free-phase. While various ways of accomplishing the required separation have been developed, as exemplified in U.S. Pat. Nos. 3,505,019; 3,555,143; 3,646,346; 3,720,760; and 3,793,445, all require at least one separate manipulative step, such as filtering, centrifuging, washing, or draining a column to insure efficient separation of the bound and free phases. Such separation is often accomplished by forming a system comprised of an insoluble portion containing the bound-phase and a liquid portion containing the free-phase such that the amount of radioactive label in either portion is a function of the extent of binding of the

labeled material, and thus a function of the amount of ligand in the sample tested. The term "heterogeneous" as generally used by the scientific community and as applied herein, means those specific binding assays wherein a separation of the bound- and free-phases is accomplished. Such a separation is necessary to carry out a specific binding assay where the labeled material in the bound-phase is indistinguishable from that in the free-phase.

Because of the hazard and difficulty of handling radioactive materials, there have been many attempts to devise convenient specific binding assay systems which are as sensitive and rapid as radioimmunoassays but which utilize features other than radioactivity as the means for monitoring the binding reaction. As will be discussed more fully hereinafter, materials which have been utilized as the labeling substance in place of radioactive atoms or molecules include such diverse materials as enzymes, fluorescent molecules, and bacteriophages.

Exemplary of methods which have been developed using an enzyme as the labeling substance are those described in U.S. Pat. Nos. 3,654,090; 3,791,932; 3,839,153; 3,850,752; and 3,879,262 and in the *Journal of Immunological Methods* 1: 247(1972) and the *Journal of Immunology* 109:129(1972). In each of the described methods an enzyme is chemically coupled to either the ligand to be detected or a binding partner thereof and an appropriate heterogeneous specific binding reaction scheme is constructed whereby after incubation with a sample, the amount of enzymatic activity associated with either the insoluble portion or the liquid portion is a function of the amount of ligand in the sample. The problems associated with the synthesis and characterization of the enzyme-conjugates are serious short comings of this approach.

Of interest is the enzyme-tagged immunoassay described in U.S. Pat. No. 3,817,837. This method does not require the use of a partitioned (i.e. insoluble portion/liquid portion) specific binding reaction system and the separation procedure necessitated thereby since the enzyme-tagged ligand is designed such that upon reaction with the binding partner of the ligand, enzymatic activity is inhibited. Thus, the ratio of bound tagged material to that in free form can be determined by monitoring changes in enzymatic activity. Nonetheless, this method suffers from the difficulty of preparing well-characterized enzyme-tagged conjugates and of finding enzymes that will fit the basic design of the system.

British Pat. No. 1,392,403 and French Pat. No. 2,201,299, which patents correspond to U.S. Pat. No. 3,880,934, describe a specific binding assay which utilizes a non-active precursor of a spectrophotometrically-active substance as the labeling substance. After incubation of the sample with the specific binding reaction system, the insoluble and liquid portions are separated and the amount of labeling substance present in the liquid portion, which is a function of the amount of ligand to be detected in the sample, is determined by carrying out reaction steps that transform the inactive labeling substance into a chromogen or fluorometrically active material which is then measured by conventional means.

Other specific binding assay methods employing different types of labeling substances are disclosed in: U.S. Pat. No. 3,850,578 which discloses the use of electron

spin resonance as a labeling means; U.S. Pat. No. 3,901,654 which discloses the use of fluorescence quenching and enhancement as a labeling means; and Report No. PB-224,875 of the National Technical Information Service (NTIS) of the U.S. Department of Commerce (1973) which describes an unsuccessful attempt to use hemin chloride as a labeling substance in a heterogeneous assay system monitored by a chemiluminescence reaction. *Nature* 219:186(1968) describes in great detail certain radioimmunoassay procedures and makes a passing reference of a very general nature to the possible use of coenzymes and viruses in place of radioisotopes as labeling substances. However, the author provides no enlightenment as to how to carry out an assay using such alternative labeling substances, or in fact as to whether such an assay would be operable. For further background, reference may be had to *Principles of Competitive Protein-Binding Assays*, ed. Odell and Daughday (J. B. Lippincott Co., Philadelphia, 1972) which discusses in breadth the various known assay schemes and the different materials and features that have been used as labels for specific binding assays.

Even though many new types of specific binding assays have been suggested and investigated, the radioimmunoassay and the various enzyme-tagged immunoassays remain the most widely used and improved. However, both types of systems have obvious shortcomings, the radioimmunoassay in its use of radioactive material which is hazardous and requires careful handling and the enzyme-tagged immunoassays in the difficulty of preparing useful enzyme-tagged conjugates.

It is therefore an object of the present invention to provide a novel method and means for detecting a ligand in a liquid which do not employ inconvenient radioactive materials or modified enzymes as the labeling substance.

Further, it is an object of the present invention to provide a heterogeneous specific binding assay method and means which are more versatile and convenient than those of the prior art.

Another object of the present invention is to provide a heterogeneous specific binding assay method and means which employ a labeling substance which is capable of being coupled to the ligand or to a specific binding partner thereof more conveniently than can the enzyme of the prior art method.

It is also an object of the present invention to provide a heterogeneous specific binding assay method and means which employ a conjugate comprising a labeling substance which is more conveniently detectable using a wide variety of sensitive monitoring reaction systems than is the enzyme in the prior art method.

#### SUMMARY OF THE INVENTION

The present invention provides a highly convenient, versatile, and sensitive improved heterogeneous specific binding assay method and means based on the use of, as labeling substance, a substance which exhibits reactant activity as a constituent of a predetermined reaction system, such substance being referred to herein as the reactant. The inventive labeling substance may be used in any of the conventional heterogeneous specific binding assay schemes. The amount of the reactant present in either of the bound- and free-phases is determined by contacting either phase with at least one reagent which forms, with the reactant, the predetermined reaction system which serves as means for monitoring the specific binding reaction. Quantitative determinations are

carried out by comparing the amount of reactant activity measured in one phase to those produced by the same assay of liquid media containing known amounts of the ligand under determination.

The improved method generally comprises the steps of (a) contacting the liquid test medium with reagent means which includes a labeled constituent comprising a conjugate of a reactant as defined herein, as labeling substance, and a binding component and which forms, with the ligand to be determined, a binding reaction system producing a bound-phase and a free-phase of said labeled constituent, the quantity of said labeling substance resulting in said bound-phase being a function of the amount of the ligand present in the test medium; (b) separating said bound-phase from said free-phase; and (c) determining the quantity of said reactant in said bound- or free-phase, and thereby the amount of the ligand in the test medium, by assessing the reactant activity therein. As will be more fully discussed herein-after, the binding reaction system may take the form of any of the known conventional techniques such as those employed in radioimmunoassay systems and in heterogeneous enzyme immunoassay systems.

The monitoring reaction system preferably is enzyme-catalyzed. Usually, a monitoring reaction system is selected which is highly sensitive to the reactant in the conjugate. Luminescent or fluorescent reaction systems are very useful in this regard. Particularly preferred are cyclic reaction systems, especially those in which the reactant is the cycled material. Of the preferred cyclic reaction systems, those which are enzyme-catalyzed are particularly advantageous. The reactant in the conjugate is usually an enzymatic reactant, such as an enzyme substrate or, as is particularly preferred, a coenzyme, and preferably has a molecular weight of less than 9000.

#### DESCRIPTION OF THE PREFERRED EMBODIMENTS

In the context of this disclosure, the following terms shall be defined as follows: ligand is the substance, or group of substances, whose presence or the amount thereof in a liquid medium is to be determined; specific binding partner of the ligand is any substance, or group of substances, which has a specific binding affinity for the ligand to the exclusion of other substances; and specific binding analog of the ligand is any substance, or group of substances; which behaves essentially the same as the ligand with respect to the binding affinity of the specific binding partner for the ligand.

The specific binding reagent means may take many different forms. In general, such means comprises three basic constituents, which are (1) the ligand to be detected, (2) a specific binding partner of the ligand, and (3) a labeled constituent which is normally a labeled form of (a) the ligand, (b) a specific binding analog of the ligand, or (c) the specific binding partner. The binding reaction constituents are combined simultaneously or in a series of additions, and with an appropriate incubation period or periods the labeled constituent becomes bound to its corresponding competing binding partners such that the extent of binding, i.e. the ratio of the amount of labeled constituent bound to a binding partner to that unbound, is a function of the amount of ligand present. To follow is a brief description of some of the different binding reaction schemes that may be used in carrying out the method of the present invention.



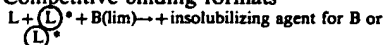
While in conventional heterogeneous specific binding assay methods, such as radioimmunoassays and heterogeneous enzyme immunoassays, the labeling characteristic in the labeled conjugate, such as radioactivity or enzymatic activity, is essentially the same for the bound- and free-forms of the conjugate, according to the present method, the activity of the reactant, as labeling substance, is in certain cases affected by binding of the labeled conjugate. In such a situation the monitoring reaction exhibits a relatively constant character where the ligand is absent from the liquid medium, or is present in an insignificantly small amount. When the ligand is present in the liquid medium, a characteristic or property of the monitoring reaction would be altered. Generally, the activity of the conjugated reactant would be the extent or rate at which the reactant is capable of participating in the monitoring reaction. Thus, the character of the monitoring reaction would be altered by the presence of the ligand in the liquid medium, usually with respect to either the aggregate reaction rate thereof or the equilibrium quantity of one or more reaction products produced thereby. Usually, in this situation, the ability of the conjugated reactant to participate in the monitoring reaction is decreased upon reaction between the specific binding substance to which it is conjugated and a specific binding counterpart of such specific binding substance, that is, the conjugate in its free state is more active in the monitoring reaction than in its bound state.

For the diagrams which are set out hereinafter, the following legend shall apply:

Symbol	Definition
$\text{L}$	ligand to be detected
$\text{L}$	ligand or specific binding analog thereof
$\text{B}$	binding partner for the ligand
$\bullet$	labeling substance, i.e. reactant
$\text{—}$	insoluble phase
$\text{—}$	incubation period followed by appropriate separation
(lim)	limited; present in an amount less than that capable of being bound to the total available binding sites under the selected reaction conditions during the selected incubation period; i.e. the constituent for which the other constituents compete for binding
(exc)	excess, present in an amount greater than that capable of being bound by the total available binding sites under the selected reaction conditions during the selected incubation period

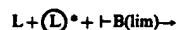
## HETEROGENEOUS ASSAY SCHEMES

### 1. Competitive binding formats

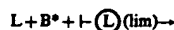


This is the classical competitive binding approach. Examples of such insolubilizing agents are specific precipitating antibodies, specific insolubilized antibodies, and, where B or  $\text{L}^*$  is a proteinaceous material, protein precipitators such as ammonium sulfate, or where B or  $\text{L}^*$  is a small adsorbable molecule, dextran-coated

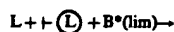
charcoal. Description of parallel systems may be found in *Biochem. J.* 88:137(1963) and U.S. Pat. No. 3,839,153.



This approach is commonly referred to as the solid-phase technique. Descriptions of parallel radioimmunoassay and enzyme immunoassay techniques may be found in U.S. Pat. Nos. 3,505,019; 3,555,143; 3,646,346; and 3,654,090.

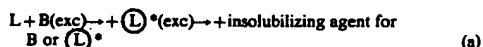


Reference: U.S. Pat. No. 3,654,090.

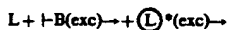


Reference: U.S. Pat. No. 3,850,752.

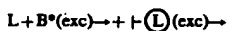
### 2. Sequential saturation formats



In the sequential saturation technique, some or all the binding sites on B remaining after the first incubation period are bound by the labeled constituent.



Descriptions of parallel radioimmunoassay and enzyme immunoassay techniques may be found in U.S. Pat. No. 3,720,760 and *J. Immunol.* 209:129(1972).



### 3. "Sandwich" format



In the sandwich technique, some or all of the ligand molecules bound to the insolubilized binding partners are bound by the labeled constituent.

Reference: U.S. Pat. No. 3,720,760.

### 4. Solid-phase dilution format



In this technique, the ligand and the labeled constituent are bound to a non-specific binder and thereafter proportional amounts are dissociated therefrom by binding with a binding partner having a greater affinity for the ligand and the labeled constituent. The most useful form of this technique employs a column of the non-specific binder as described in U.S. Pat. No. 3,659,104. Such a technique is useful where the ligand is bound to endogenous binding substances in the sample which unless removed would interfere with the competitive binding reaction. Upon being bound to the non-specific binder, the endogenous binding substances may be removed by appropriate washes.

For further discussion of the parameters involved in conventional heterogeneous assay systems, such as more detailed descriptions of assay formats and alternative separation techniques, reference may be had to *Principles of Competitive Protein-Binding Assays*, ed. Odell and Daughaday (J. B. Lippincott Co., Philadelphia, 1972).

It is contemplated that manipulative schemes involving other orders of addition and other binding reaction

formats may be devised for carrying out heterogeneous specific binding assays without departing from the inventive concept embodied herein.

The step of assessing the activity of the conjugated reactant as a constituent of the predetermined monitoring reaction system in either of the bound- or free-phase with at least one substance which forms with the conjugated reactant, the monitoring reaction, and measuring a characteristic of such reaction. The monitoring reaction system may comprise a single chemical transformation or a plurality of series of chemical transformations.

Where an enzyme-catalyzed reaction system is used, it includes, in addition to the conjugated reactant, at least one enzyme and may include one or more enzymatic reactants such as substrates and coenzymes. Such enzyme-catalyzed reaction system may comprise a single simple enzymatic reaction or a complex series of enzymatic and non-enzymatic reactions. For instance, the enzyme-catalyzed reaction system may consist of a single enzyme-catalyzed degradation or dissociation reaction. In such a system, the conjugated reactant is the enzyme substrate which undergoes degradation or dissociation, and the only component of the reaction system necessary to be contacted with the selected bound- or free-phase is an enzyme which catalyzes the degradation or dissociation reaction. A more complex enzyme-catalyzed reaction system may consist of a single enzymatic reaction involving two or more reactants or may consist of a series of reactions involving several reactions, at least one of which reactions is enzyme-catalyzed. In such a system, the conjugated reactant would be one of the enzymatic reactants in the enzyme-catalyzed reaction and the selected bound- or free-phase would be contacted with the appropriate enzyme and reactant constituents, other than that in the conjugate, necessary to provide the selected enzyme-catalyzed reaction system.

It is further contemplated that the enzyme-catalyzed reaction system may comprise a biochemical system as complex as the metabolic system of a biological cell such as a microorganism. For example, a nutrient substance essential to the growth of a particular microorganism may be selected as the reactant in the conjugate. Reactant activity would be measurably by monitoring a characteristic of the microorganism, such as the rate of microorganism growth, when such microorganism would be placed in an environment wherein the only source of the reactant nutrient substance is the conjugate.

The appropriate reaction constituents which form, together with the reactant in the conjugate, the monitoring reaction system may be contacted with the selected separated phase mixture singularly or in any combination either prior to, simultaneous with, or subsequent to initiation of the specific binding reaction. After initiation of the specific binding reaction, the reaction mixture, which may include any or all of the necessary components for the monitoring reaction is usually incubated for a predetermined period or periods of time before separation of the resulting bound- and free-phases. After separation, any components which are necessary for the monitoring reaction and which are not already present in sufficient quantities in the selected separated phase are added thereto, and reactant activity therein is assessed as an indication of the pressure or amount of the ligand in the liquid medium.

When the reaction rate of the monitoring reaction is the characteristic used to assess reactant activity in the

selected bound- or free-phase, as is preferred, such rate is usually determined by measuring the rate of disappearance of a reactant or the rate of appearance of a reaction product. Such measurement can be accomplished by a wide variety of methods including the conventional chromatographic, gravimetric, potentiometric, spectrophotometric, fluorometric, turbidimetric, and volumetric analysis techniques. Since the present method is primarily designed for the detection of low concentrations of ligands, highly sensitive reaction systems have been developed for use in conjunction with the novel specific binding reaction system.

One preferred form of the monitoring reaction includes a luminescent reaction system, preferably enzyme-catalyzed, such as a reaction exhibiting the phenomenon of bioluminescence or chemiluminescence. The reactant in the conjugate may be a reactant in either the light-producing reaction or a reaction which is preliminary to an enzymatic or nonenzymatic luminescent reaction. The activity of the conjugated reactant can be assessed by following the rate of light production or the total amount, peak intensity, or character of the light produced. Examples of luminescent reaction systems are given in Table A in which the following abbreviations are used:

ATP: adenosine triphosphate  
AMP: adenosine monophosphate  
NAD: nicotinamide adenine dinucleotide  
NADH: reduced nicotinamide adenine dinucleotide  
FMN: flavin mononucleotide  
FMNH<sub>2</sub>: reduced flavin mononucleotide  
hν: electromagnetic radiation, usually in the infrared, visible, or ultraviolet region

TABLE A

Luminescent Reaction System	Conjugated Reactant
A. ATP + reduced luciferin $\xrightarrow[\text{(fire fly)}]{\text{luciferase}}$	ATP or reduced luciferin
hν + AMP + oxidized luciferin	luciferin
B. FMNH <sub>2</sub> + long-chain aldehyde + O <sub>2</sub> $\xrightarrow[\text{(P. fisheri)}]{\text{luciferase}}$	FMNH <sub>2</sub> or long-chain aldehyde
hν + FMN + long-chain acid + H <sub>2</sub> O	
C. (1) NADH + FMN + H <sup>+</sup> $\xrightarrow{\text{NADH dehydrogenase}}$	NADH or FMN
NAD + FMNH <sub>2</sub>	
(2) FMNH <sub>2</sub> + long-chain aldehyde + O <sub>2</sub> $\xrightarrow[\text{(P. fisheri)}]{\text{luciferase}}$	
hν + FMN + long-chain acid + H <sub>2</sub> O	
D. (1) 3',5'-adenosine diphosphate + reduced luciferin sulfate $\xrightarrow{\text{transferase}}$	3',5'-adenosine diphosphate or reduced luciferin
adenosine-3'-phosphate-5'-phosphosulfate + reduced luciferin	
(2) reduced luciferin + O <sub>2</sub> $\rightarrow$	
hν + oxidized luciferin	
E. luminol + H <sub>2</sub> O <sub>2</sub> $\xrightarrow{\text{peroxidase*}}$	luminol
hν + aminophthalate + N <sub>2</sub>	
F. reduced pyrogallol + H <sub>2</sub> O <sub>2</sub> $\xrightarrow{\text{peroxidase*}}$	reduced pyrogallol
hν + oxidized pyrogallol + H <sub>2</sub> O	
G. luminol + O <sub>2</sub> $\xrightarrow{\text{oxygenase}}$	luminol
hν + aminophthalate + N <sub>2</sub>	
H. reduced pyrogallol + O <sub>2</sub> $\xrightarrow{\text{oxygenase}}$	reduced pyrogallol
hν + oxidized pyrogallol	

TABLE A-continued

Luminescent Reaction System	Conjugated Reactant
I. isoluminol + H <sub>2</sub> O <sub>2</sub> lactoperoxidase → hv + aminophthalate + N <sub>2</sub>	isoluminol
J. isoluminol + KO <sub>2</sub> → hv + aminophthalate + N <sub>2</sub>	isoluminol

\*or catalase

Further details and discussion concerning luminescent reaction systems which may be used in the present method may be found in the following references:

*J. Biol. Chem.* 236:48(1961)

*J. Amer. Chem. Soc.* 89:3944(1967).

Cornier et al *Bioluminescence in Progress*, ed. Johnson et al, Princeton University Press (New Jersey, 1966) pp. 363-84.

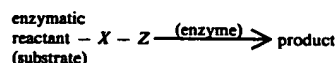
Kries, P. *Purification and Properties of Renilla Luciferase*, doctoral thesis University of Georgia (1967).

*Am. J. Physiol.* 41:454(1961).

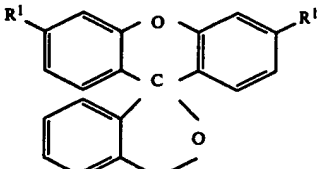
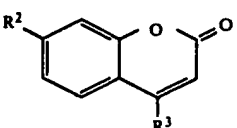
*Biol. Bull.* 51:89(1926).

*J. Biol. Chem.* 243:4714(1968).

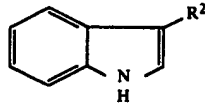
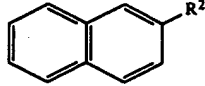
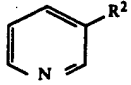
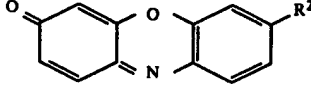
Another type of preferred, sensitive, monitoring reaction involves the phenomenon of fluorescence and is enzyme-catalyzed. In such a reaction system the reactant in the conjugate is a substrate in an enzymatic reaction which produces a product which has a fluorescent property which distinguishes it from the conjugated substrate. A general reaction scheme for such an enzyme-catalyzed reaction system is as follows:



wherein X is an enzyme-cleavable bond or linking group, such as an ester or amido group, and Z is a specific binding substance which, depending upon the specific binding reaction technique used, is the ligand, a specific binding analog of the ligand, or a specific binding partner of the ligand. Specific conjugates which may be used in this type of reaction system are various enzyme-cleavable modifications and derivatives of fluorescein, umbelliferone, 3-indole,  $\beta$ -naphthol, 3-pyridol, resorufin, and so forth. Examples of possible structural formulas of such derivatives are as follows:

Derivative	Formula
fluorescein	
umbelliferone	

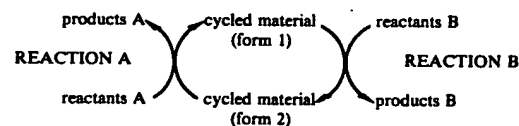
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Derivative	Formula
3-indole	
$\beta$ -naphthol	
3-pyridol	
resorufin	

wherein R<sup>1</sup> is —OH or X—Z (as defined above in this paragraph), R<sup>2</sup> is —X—Z, and R<sup>3</sup> is —H or —CH<sub>3</sub>.

A reaction system which is particularly preferably for use in assessing the activity of the conjugated reactant in the selected separated phase is a cyclic or cycling reaction system. Such a reaction system is one in which a product of a first reaction is a reactant in a second reaction, which second reaction has as one of its products a substance that is also a reactant in the first reaction.

The following diagram illustrates a model of a cyclic reaction system:



In the above model cyclic reaction system, a small amount of cycled material, if provided with sufficient amounts of reactants A and B, will generate large amounts of products A and B. Since the rate and amount of product produced by the reactions constituting the cyclic reaction system is highly sensitive to the amount of cycled material present, it is most preferred to use the cycled material as the reactant in the conjugate of the present invention. Examples of cycling reaction systems contemplated for use in conjunction with the novel specific binding reaction system of the present invention are given in Tables B, C, and D.

TABLE B

reaction	reactant A or product B	enzyme	product A or reactant B
1	lactaldehyde	alcohol dehydrogenase	propanediol
2	$\alpha$ -ketoglutarate + NH <sub>3</sub>	glutamic dehydrogenase	glutamate
3	oxaloacetate	malic dehydrogenase	malate
4	acetaldehyde	alcohol dehydrogenase	ethanol

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TABLE B-continued

	product A	NAD <sup>+</sup>	reactant B
	enzyme	enzyme	
	reactant A	NADH <sup>+</sup>	product B
	reactant A or product B		reactant B or product A
reaction	product B	enzyme	product A
5	α-ketoglutarate + CO <sub>2</sub>	isocitric dehydrogenase	isocitrate
6	dihydroxyacetone phosphate	α-glycerol phosphate dehydrogenase	L-α-glycerol phosphate
7	pyruvate	lactic dehydrogenase	lactate
8	1,3-diphosphoglycerate	glyceraldehyde-3-phosphate dehydrogenase	glyceraldehyde-3-phosphate + phosphate

\*nicotinamide adenine dinucleotide  
\*\*reduced NAD

TABLE C

	product A	NADP <sup>+</sup>	reactant B
	enzyme	enzyme	
	reactant A	NADPH <sup>+</sup>	product B
	reactant A or product B		reactant B or product A
reaction	product B	enzyme	product A
1	6-phosphogluconate	glucose-6-phosphate dehydrogenase	glucose-6-phosphate
2	oxidized glutathione	glutathione reductase	reduced glutathione
3	p-benzoquinone	quinone reductase	hydroquinone
4	nitrate	nitrate reductase	nitrite
5	α-ketoglutarate + NH <sub>3</sub>	glutamic dehydrogenase	glutamate

\*nicotinamide adenine dinucleotide phosphate  
\*\*reduced NADP

It should be noted that the cyclic reaction systems illustrated in Tables B and C comprise the combination of any one of the reactions listed in the respective tables with any other reaction listed therein. For example, reaction 1 in Table B may be paired with any one of reactions 2-8 to form a useful cyclic reaction system. Thus, Tables B and C represent respectively 56 and 20 possible cyclic reaction systems for use in the present invention.

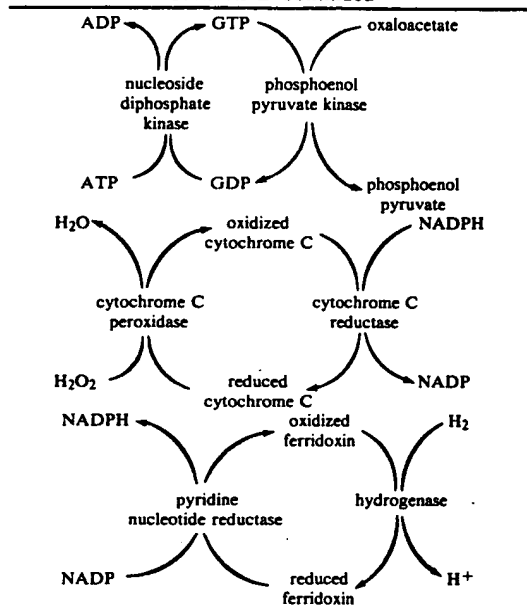
In addition to the cyclic reaction systems represented in Tables B and C, it is contemplated that one of the reactions in the cyclic reaction system may involve the enzymatic or non-enzymatic conversion of a spectrophotometric indicator, preferably colorimetric. An example of a cyclic reaction system involving a conversion of an indicator is the system produced by combining one of the reactant B-product B reactions from Table B with a reaction comprising an oxidation-reduction indicator and an electron transfer agent. As electron transfer agent, phenazinemethosulfate may be used. Useful indicators include the oxidized forms of nitrotetrazolium, thiazolyl blue, and dichlorophenoldophenol.

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TABLE D

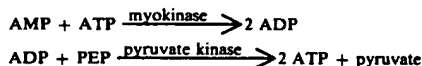
5	reduced cytochrome C	FMN <sup>a</sup>	NADPH
	cytochrome C reductase	cytochrome C reductase	
10	cytochrome C	FMNH <sub>2</sub> <sup>b</sup>	NADP
	<sup>a</sup> flavin mononucleotide		
	<sup>b</sup> reduced FMN		
15	H <sub>2</sub> O <sub>2</sub>	FAD <sup>c</sup>	D-aminoacid
	D-aminoacid oxidase	D-aminoacid oxidase	
20	O <sub>2</sub>	FADH <sub>2</sub> <sup>d</sup>	α-ketoacid + NH <sub>3</sub>
	<sup>c</sup> flavin adenine dinucleotide		
	<sup>d</sup> reduced FAD		
25	α-ketoglutarate	α-aminoacid	O <sub>2</sub>
	transaminase	L-aminoacid oxidase	
30	glutamate	α-ketoacid	H <sub>2</sub> O <sub>2</sub>
	phosphate	ADP <sup>e</sup>	phosphoenolpyruvate
	adenosine triphosphatase	pyruvate kinase	
35		ATP <sup>f</sup>	pyruvate
	<sup>e</sup> adenosine diphosphate / adenosine triphosphate		
40	succinate + GTP <sup>g</sup>	coenzyme A	α-ketoglutarate + NAD
	succinic thiokinase	α-ketoglutarate dehydrogenase	
45	phosphate + GDP <sup>h</sup>	succinyl-coenzyme A	NADH + CO <sub>2</sub>
	<sup>g</sup> guanosine triphosphate		
	<sup>h</sup> guanosine diphosphate		
50	ascorbate	oxidized glutathione	NADPH
	dehydroascorbate reductase	glutathione reductase	
55	dehydroascorbate	reduced glutathione	NADP
	oxidized glutathione	ascorbate	O <sub>2</sub>
60	dehydroascorbate reductase	ascorbate oxidase	H <sub>2</sub> O
	reduced glutathione	dehydroascorbate	

TABLE D-continued



In forming any of the cyclic reaction systems illustrated in Tables B, C, and D, where a component in the reaction system is in an ionic form, it may of course be added in a salt or acid form which is ionizable upon contacting the liquid medium. A water soluble salt or acid of such component is usually preferred.

It is also contemplated that an exponential cyclic reaction system may be included in the monitoring reaction system. An example of an exponential cyclic reaction system is as follows:



Such a cyclic reaction is autocatalytic in the sense that during each cycle the amount of cycled material is doubled. The cycling rate therefore increases exponentially with time and affords a high degree of sensitivity. Further details and discussion relating to such cyclic reactions may be found in *J. Biol. Chem.* 247:3558-70(1972).

Where a cyclic reaction system is used as a means of assessing any change in activity of the conjugated reactant, the rate of disappearance of a reactant or rate of appearance of a reaction product can be determined by conventional techniques or by using one or more additional cycling systems following by a conventional determination of the aggregate reaction rate.

The use of a cyclic reaction system in conjunction with the heterogeneous specific binding reaction system provides a high degree of assay versatility as well as sensitivity. A single reactant-specific binding substance conjugate may be used with a multiplicity of reactions to form cyclic systems which have sensitivities varying over a wide range and which provide a wide variety of responses detectable by the senses or artificial means. Such versatility is lacking in the prior art.

The present invention may be applied to the detection of any ligand for which there is a specific binding part-

ner. The ligand usually is a peptide, protein, carbohydrate, glycoprotein, steroid, or other organic molecule for which a specific binding partner exists in biological systems or can be synthesized. The ligand, in functional terms, is usually selected from the group consisting of antigens and antibodies thereto; haptens and antibodies thereto; and hormones, vitamins, metabolites and pharmacological agents, and their receptors and binding substances. Specific examples of ligands which may be detected using the present invention are hormones such as insulin, chorionic gonadotropin, thyroxine, liothyronine, and estriol; antigens and haptens such as ferritin, bradykinin, prostaglandins, and tumor specific antigens; vitamins such as biotin, vitamin B<sub>12</sub>, folic acid, vitamin E, and ascorbic acid; metabolites such as 3',5' adenosine monophosphate and 3',5' guanosine monophosphate; pharmacological agents such as dilantin, digoxin, morphine, digitoxin, and barbiturates; antibodies such as microsomal antibody and antibodies to hepatitis and allergens; and specific binding receptors such as thyroxine binding globulin, avidin, intrinsic factor, and transcobalamin.

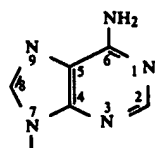
In the conjugate of the present invention, the reactant is coupled or bound to a specific binding substance, which is the ligand, a specific binding analog of the ligand, or a specific binding partner of the ligand, depending upon the assay scheme selected, such that a measurable amount of activity of the reactant is retained. The bond between the reactant and the specific binding substance is usually substantially irreversible under the conditions of the assay such as where the predetermined monitoring reaction in which the reactant has activity is not designed to chemically destroy such bond as in the above-mentioned luminescent and cyclic reaction systems. However, in certain instances such bond is by design destroyed or otherwise affected by the selected monitoring reaction as a means for assessing in reactant activity. Such a case is the enzymatic fluorescent substrate reaction systems referred to previously herein.

The reactant may be directly coupled to the specific binding substance so that the molecular weight of the conjugate is less than or equal to the aggregate molecular weight of the reactant and the specific binding substance. Usually, however, the reactant and the specific binding substance are linked by a bridge group comprising between 1 and 50, and preferably between 1 and 10, carbon atoms or heteroatoms such as nitrogen, oxygen, sulfur, phosphorus and so forth. Examples of a bridge group comprising a single atom would be a methylene group (one carbon atom) and an amino group (one heteroatom). The bridge group usually has a molecular weight not exceeding 1000 and preferably less than 200. The bridge group comprises a chain of carbon atoms or heteroatoms, or a combination of both, and is joined to the reactant and the specific binding substance, or active derivative thereof, by a connecting group usually in the form of an ester, amido, ether, thioester, thioether, acetal, methylene, or amino group.

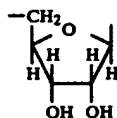
The reactant in the conjugate of the present invention may be any substance which has given (i.e. fixed or known) reactant activity as a constituent of a predetermined monitoring reaction. More particularly, for the purposes of this disclosure, the terms "reactant" and "substance having reactant activity" refer to any chemical substance which is capable of undergoing a finite measurable chemical transformation which yields one



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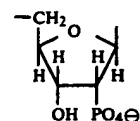


Also, the following abbreviations are used;  
Rib refers to the ribose moiety, i.e.,



Rib' refers to the phosphorylated ribose moiety, i.e.,

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Ph refers to a phosphate group;

AP derivatives refers to derivatives of adenosine-5'-phosphate, i.e., the mono- (AMP), di- (ADP), or tri- (ATP) phosphate form;

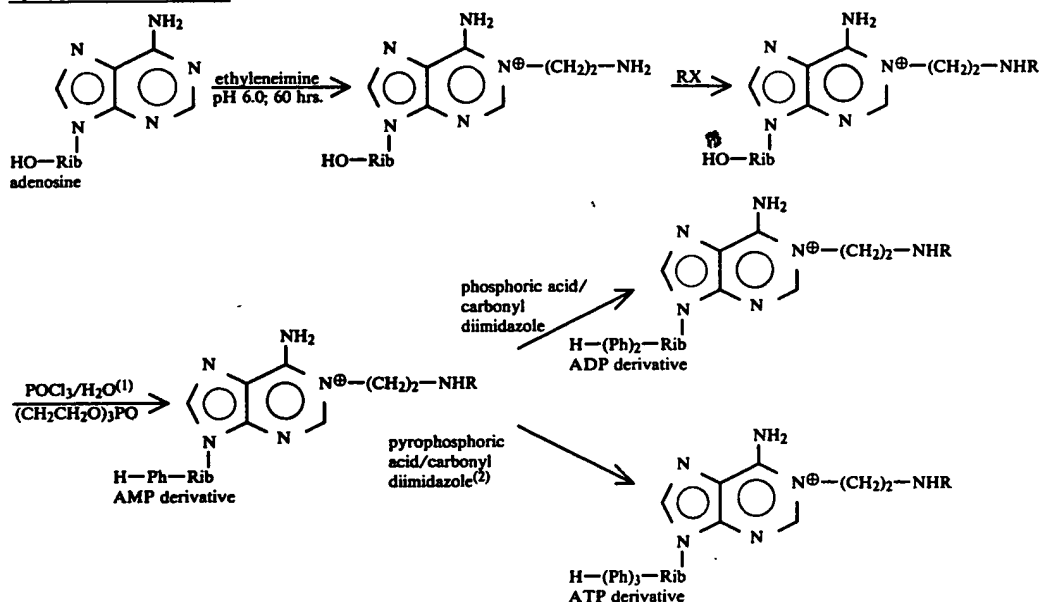
NAD derivative refers to a derivative of either nicotinamide adenine dinucleotide or a reduced form thereof;

NADP derivative refers to a derivative of either nicotinamide adenine dinucleotide phosphate or a reduced form thereof;

R refers to the specific binding substance or a modification thereof; and

X refers to a leaving group, usually a halogen.

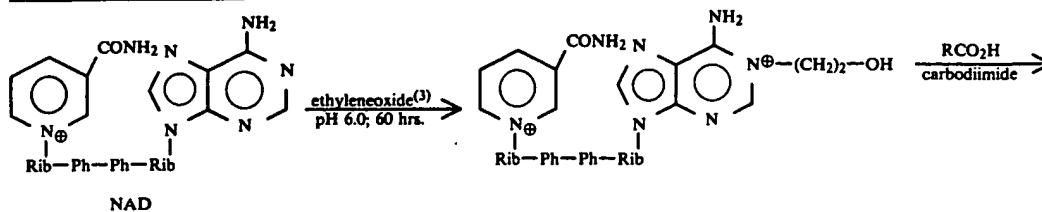
#### 1-position derivatives of AP



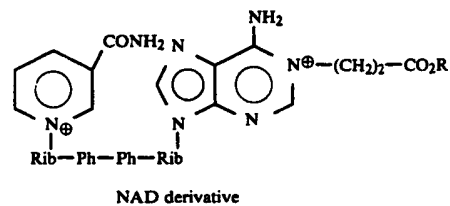
<sup>(1)</sup>Guilford, H., et al., *Chemica Scripta* 2:165 (1972).

<sup>(2)</sup>Trayer, I. P., et al., *Biochem. J.* 139:609 (1974).

#### 1-position derivative of NAD

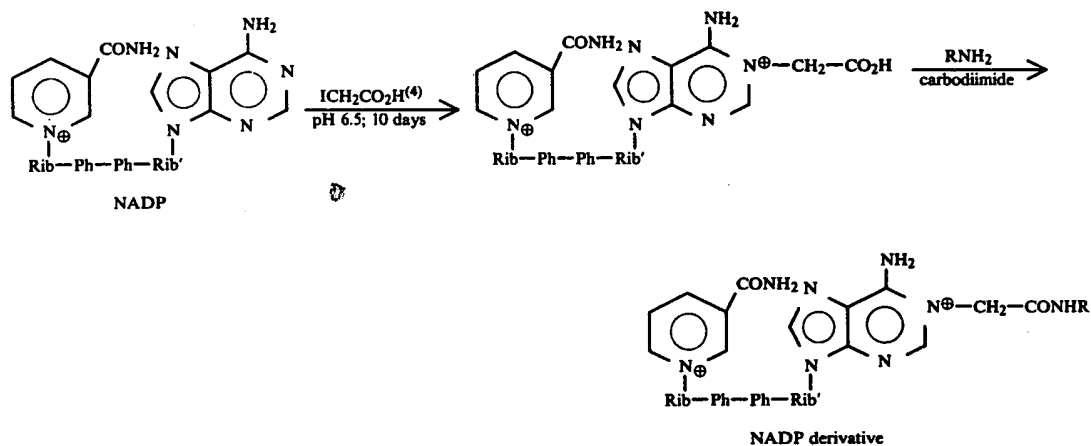


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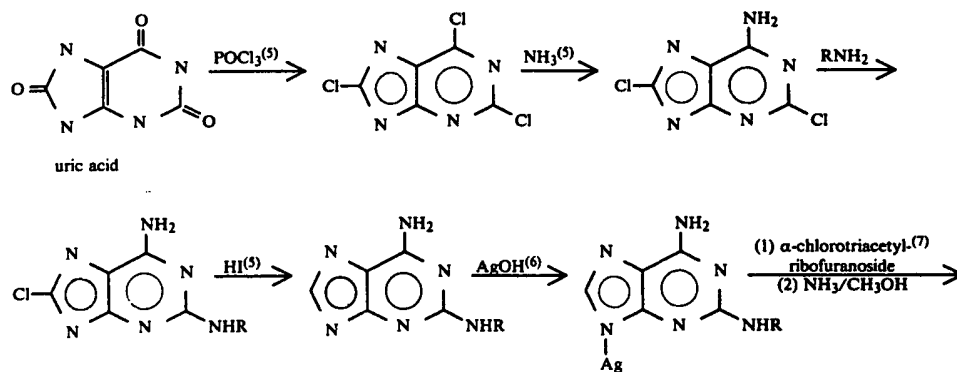
<sup>(3)</sup>Windmueller, H. G., and Kaplan, N. O., J. Biol. Chem. 236:2716 (1961).

#### 1-position derivative of NADP

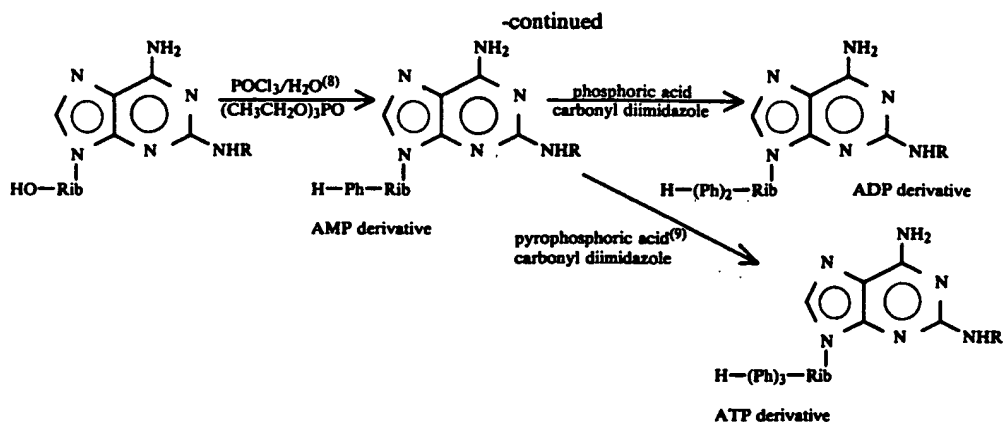


<sup>(4)</sup>Lowe, C. R. and Mosbach, K., Eur. J. Biochem. 49:511 (1974).

#### 2-position derivatives of AP







(8) Acheson, R. M., An Introduction to the Chemistry of Heterocyclic Compounds, Interscience Publ. (New York 1962), p. 308.

(6) Fischer, E., Ber. 30:2239 (1897).

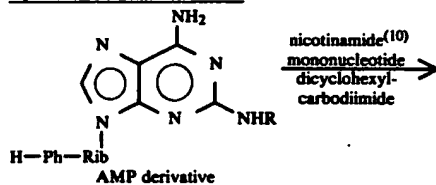
(7) Davoll et al., J. Chem. Soc., 967 (1948).

(8) Guilford, H., et al., supra.

(9) Trayer, I. P., et al., supra.

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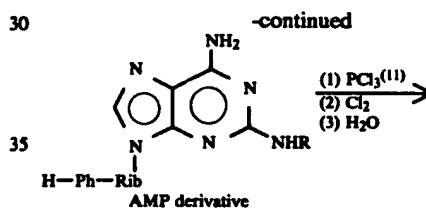
#### 2-position derivative of NAD



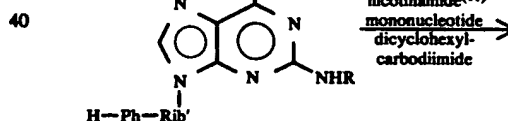
(10) Hughes, N. A., et al., J. Chem. Soc., 3733 (1957).

#### 2-position derivative of NADP

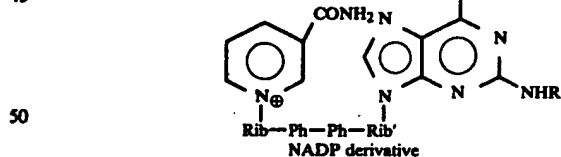
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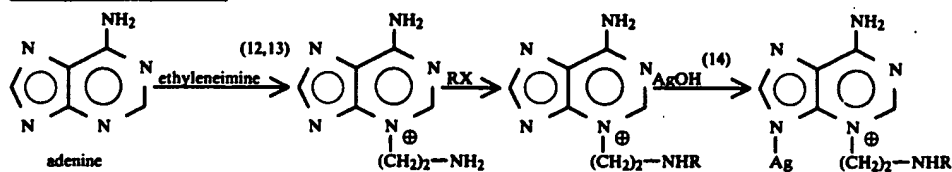


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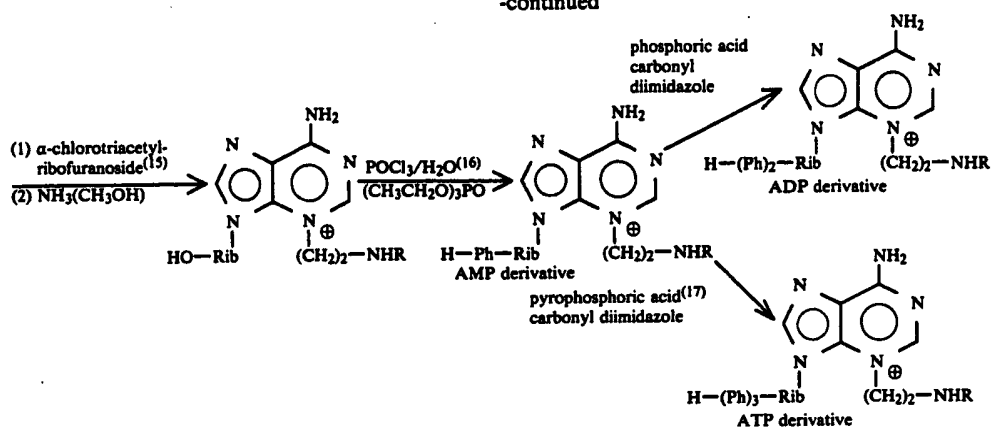
(11) Hughes, N. A., et al., supra.

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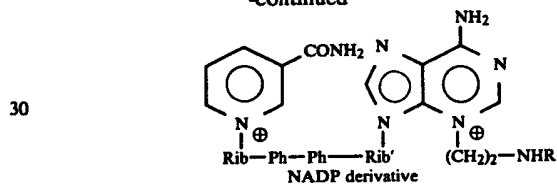
#### 3-position derivatives of AP



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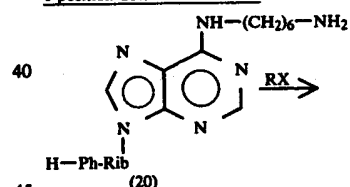
(12) Lister, J. H., in *Advances in Heterocyclic Chemistry*, ed. Kabritsky et al., Academic Press (New York, 1966), p.33.(13) Leonard, N. J., and Fujii, T. J., *J. Amer. Chem. Soc.* 35:3719 (1963).(14) Fischer, E., *supra*.(15) Davoll et al., *supra*.(16) Guilford, H., et al., *supra*.(17) Trayer, I. P., et al., *supra*.

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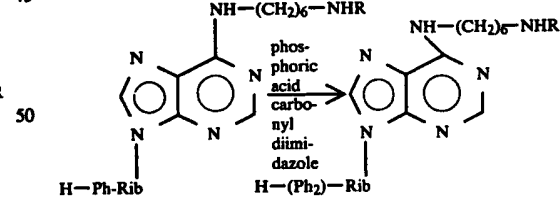


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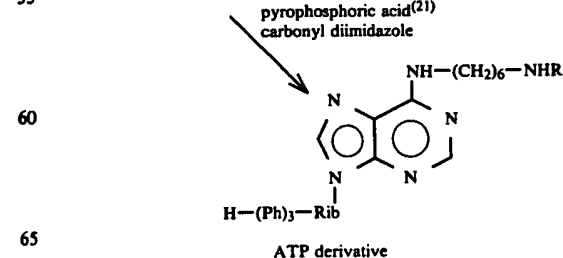
6-position derivatives of AP



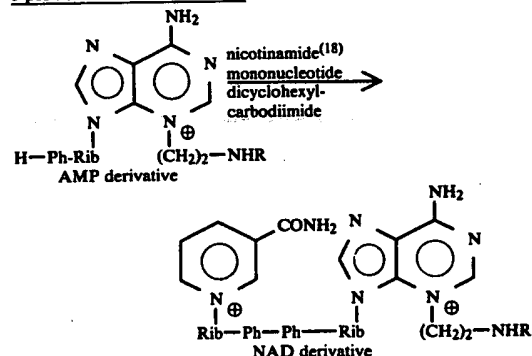
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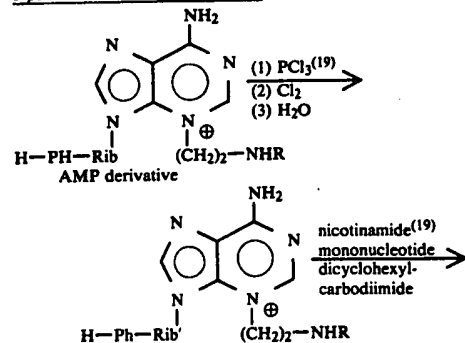
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(20) Guilford, H., et al., *supra*.(21) Trayer, I. P., et al., *supra*.

3-position derivative of NAD

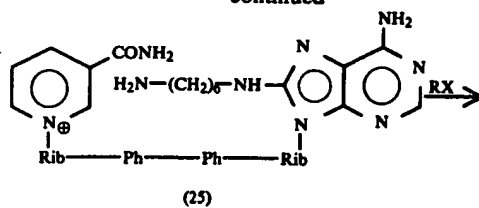
(18) Hughes, N. A., et al., *supra*.

3-position derivative of NADP



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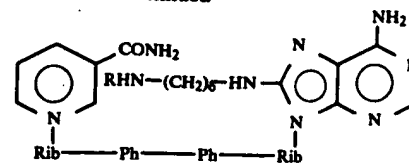
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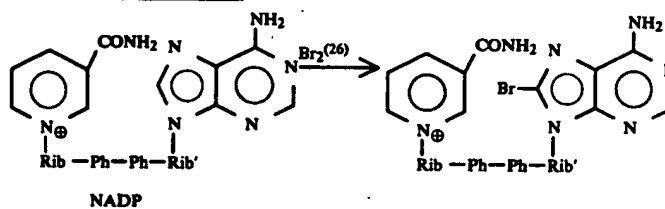
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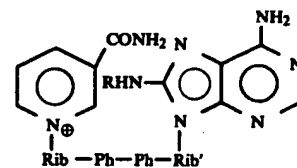
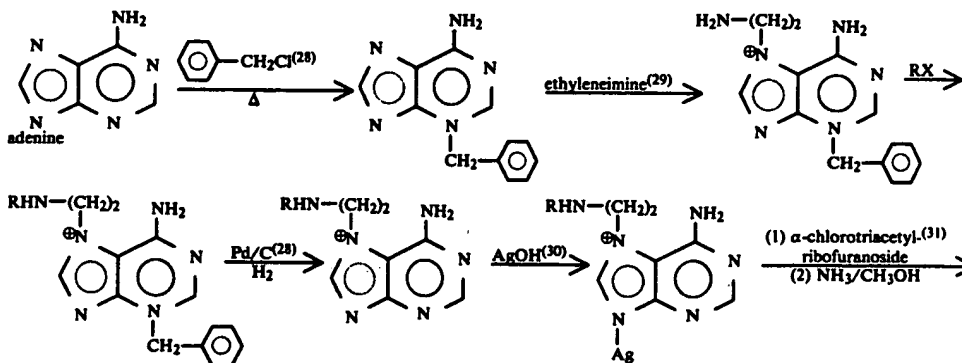
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(25) Lee, C-Y, et al., Arch. Biochem. Biophys. 163:561 (1974).

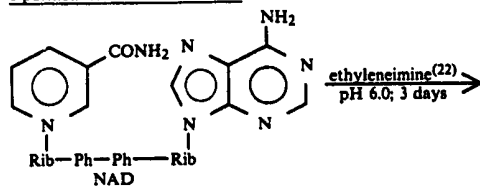
NAD derivative

8-position derivative of NADP

- (1) glucose-6-phosphate<sup>(27)</sup>  
dehydrogenase  
(2) RNH<sub>2</sub>; Δ  
(3) glutamate  
dehydrogenase

(26) Lee, C-Y, et al., *supra*.(27) Lowe, C.R. and Mosbach, R., *supra*.9-position derivatives of AP

## 6-position derivative of NAD

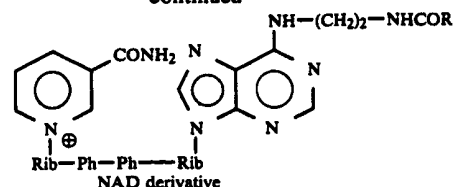


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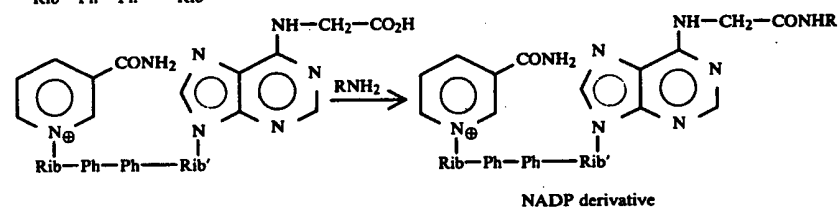
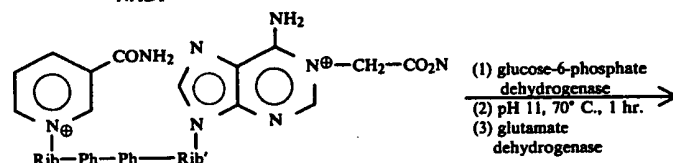
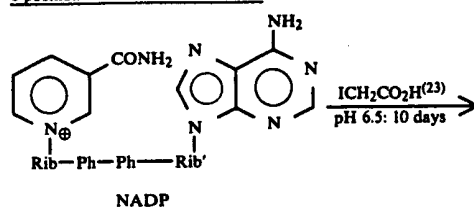
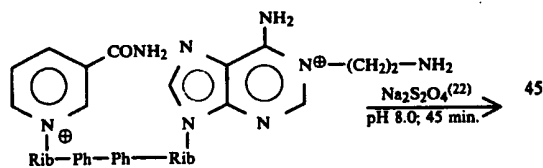
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<sup>(22)</sup>Windmueller, H. G., and Kaplan, N. O., J. Biol. Chem. 236:2716 (1961).

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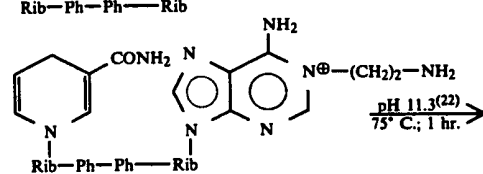


## 6-position derivative of NADP

<sup>(23)</sup>Lowe, C.R., and Mosbach, K., *supra*.

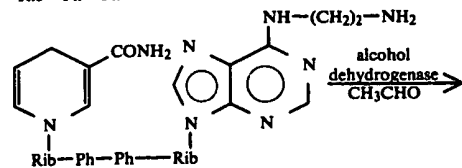
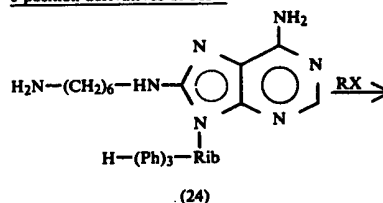
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## 8-position derivatives of AP

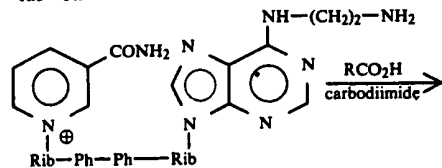


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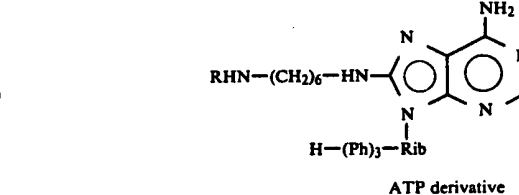
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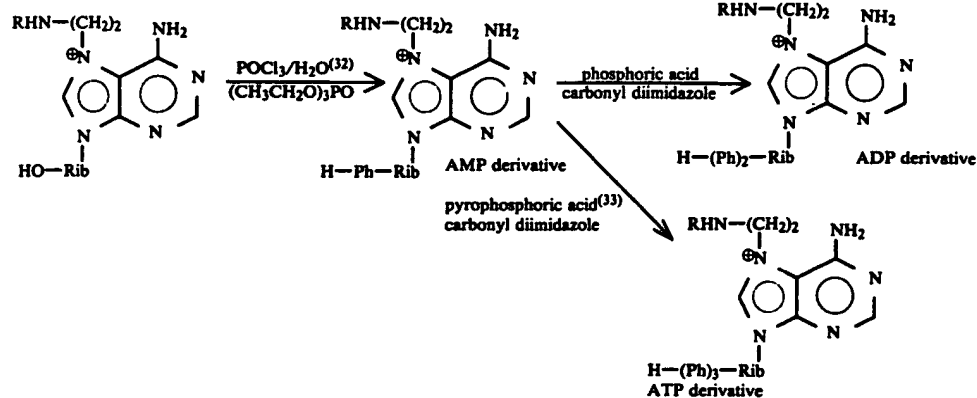
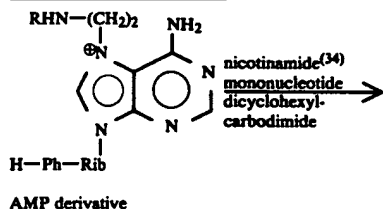
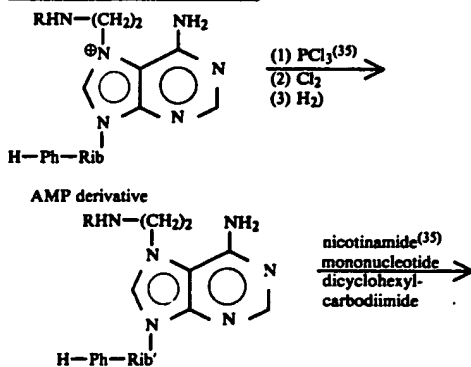


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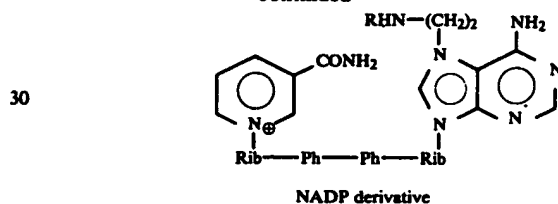
(24) Trayer, I. P., et al., *supra*.

## 8-position derivative of NAD

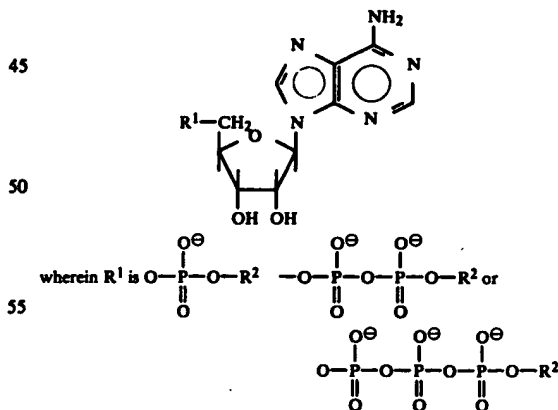
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<sup>(32)</sup>Leonard, N. J., and Fujii, T. J., *supra*.<sup>(29)</sup>Lister, J. H., *supra*.<sup>(30)</sup>Fiacher, E., *supra*.<sup>(31)</sup>Davoll, et al., *supra*.<sup>(32)</sup>Guildford, H., et al., *supra*.<sup>(33)</sup>Trayer, I. P., et al., *supra*.9-position derivative of NAD<sup>(34)</sup>Hughes, N. A., et al., *supra*.9-position derivative of NADP

-continued

<sup>(35)</sup>Hughes, N. A., et al., *supra*.

In addition to the compounds mentioned above, useful coenzyme-active conjugates include the adenosine phosphates to which are coupled the specific binding substance through the phosphate grouping. Such compounds have the following general formula:



wherein R<sup>1</sup> is  $\text{O}-\text{P}(=\text{O})(\text{O}^-)-\text{O}-\text{R}^2$  or  $\text{O}-\text{P}(=\text{O})(\text{O}^-)-\text{O}-\text{P}(=\text{O})(\text{O}^-)-\text{O}-\text{R}^2$  or  $\text{O}-\text{P}(=\text{O})(\text{O}^-)-\text{O}-\text{P}(=\text{O})(\text{O}^-)-\text{O}-\text{P}(=\text{O})(\text{O}^-)-\text{O}-\text{R}^2$

wherein R<sup>2</sup> is  $-\text{Y}-\text{Z}$ ; wherein Y is a bond or a bridge group; and wherein Z is a ligand, a specific binding analog of a ligand, or a specific binding partner of a ligand. Also, the protonized or acid forms, as well as the salt forms where appropriate, may be used.

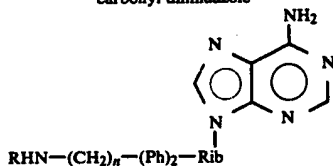
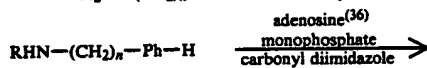
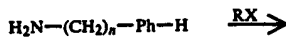
Synthesis of such compounds may be accomplished in a variety of ways. It is contemplated that the synthesis routes which are schematically illustrated below are

advantageously followed in the preparation of the useful compounds. The abbreviations used hereinbefore also apply to the illustration to follow.

#### derivatives of AP



$n = 1-10$

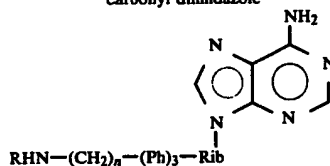
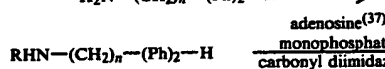
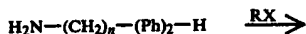


ADP derivative

<sup>(36)</sup>Trayer, I.P., et al., *Biochem. J.* 139:609 (1974).



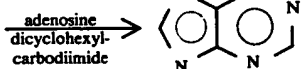
$n = 1-10$



ATP derivative



$n = 1-10$



AMP derivative

<sup>(37)</sup>Trayer, I.P., et al., *supra*.

In one form of the present invention, the components of the specific binding reaction which are to be combined with the liquid medium suspected of containing the ligand are in a liquid or solid form. The assay method may be carried out in a standard laboratory vessel such as a test tube with the specific binding reaction components and the components of the reaction system being added thereto in solid or liquid form.

It is also contemplated that one or more of the specific binding reaction components and/or one or more of the components of the monitoring reaction may be incorporated with a carrier. In one aspect, the carrier may be a liquid-holding vessel such as a test tube or capsule containing such component or components in an interior portion thereof, for instance, in the form of a liquid or loose solid or a coating on an interior surface of the vessel. In another aspect, the carrier may be in the form of matrix which is insoluble and porous, and preferably absorbent, relative to the liquid medium to be

tested. Such matrix may be in the form of bibulous papers; polymeric films, membranes, fleeces, or blocks; gels; and so forth. In such a form, the device would provide a convenient means for contacting the liquid medium to be tested, for carrying out the specific binding reaction and/or the monitoring reaction, for effecting the necessary separation, and for observing the resulting response.

The liquid medium to be tested may be a naturally occurring or artificially formed liquid suspected of containing or known to contain the ligand, and usually is a biological fluid or a liquid resulting from a dilution or other treatment thereof. Biological fluids which may be assayed following the present method include serum, plasma, urine, and amniotic, cerebral, and spinal fluids. Other materials such as solid matter, for example tissue, or gases may be assayed by reducing them to a liquid form such as by dissolution of the solid or gas in a liquid or by liquid extraction of the solid.

In contrast to the prior art assay systems, biological fluids containing substances which have reactant activity similar or identical to that of the conjugated labeling substance may be assayed for the ligand without background interference. Endogenous background reactant activity can be readily eliminated in several manners. The biological fluid can be treated to selectively destroy the endogenous reactant activity. Such treatment may involve the action of a clearing agent which chemically destroys the endogeneous activity followed by treatment to inactivate the destructive action of such clearing agent.

For instance, reactant-degrading enzymes often appear naturally in biological fluids, particularly if the reactant is a coenzyme such as NAD, NADP, or ATP. There are many inhibitors of such coenzyme-degrading enzymes, for example chelating agents which operate to deprive the enzymes of essential metal ion activators. As a specific example, such degrading enzymes are found in normal serum and have sufficient enzymatic activity to remove essentially all endogenous NAD activity from isolated serum within a few hours. The degrading activity of such enzymes may be effectively inhibited by addition of a chelating agent such as ethylene-diamine tetraacetic acid. Elimination of the degrading activity may also be accomplished by adding a specific enzyme inhibitor. For example, ATP-degrading enzymes may be inhibited by addition of  $\beta\gamma$  methyleneATP or  $\alpha\beta$  methylene ATP.

The present invention will now be illustrated, but is not intended to be limited, by the following Examples.

#### EXAMPLE 1

##### Preparation of nicotinamide 6-(2-aminoethylamino) purine dinucleotide

Two (2) grams of nicotinamide adenine dinucleotide (NAD) were dissolved in 10 ml of water and 0.6 ml of ethyleneimine was added dropwise, the pH being maintained below by the addition of 1 M perchloric acid. When addition of ethyleneimine was complete, the pH was adjusted to 4.5 and the reaction was incubated at 20°-25° C. At 24 hour intervals 0.6 ml of ethyleneimine was added and the pH readjusted to 4.5. After 96 hours, the solution was poured into 10 volumes of acetone at -10° C. The oil which formed was collected, washed with ether, and dissolved in approximately 50 ml of water in a flask.

The resulting solution was adjusted to pH 7.0-7.5 with 1 N sodium hydroxide, and 1 gram of sodium bicarbonate was added. Nitrogen was bubbled through the solution for from 4 to 5 minutes and 1 gram of sodium hydrosulfite was added. The flask was sealed tightly and allowed to stand at room temperature for 45 minutes. The solution was then oxygenated for 15 minutes and adjusted to pH 11.3 with sodium hydroxide. The solution was heated at 75° C. for 1 hour. Then the reaction mixture was cooled to room temperature and 0.6 grams of tris-(hydroxymethyl)-aminomethane was added, followed by 5 N hydrochloric acid to adjust the pH to 7.5. To the resulting solution was added 1000 International units of alcohol dehydrogenase and 1 ml of acetaldehyde. The decreasing optical density of the reaction mixture was monitored at 340 nm and when no further decrease was observed, the pH was adjusted to 3.5. The solution was poured into 10 volumes of acetone at -10° C. The oil which formed was separated and washed with ether, after which it was dissolved in 10 to 15 ml of water.

The resulting solution was introduced into a 2.5×90 cm column of Sephadex G-10, available from Pharmacia AB, Uppsala, Sweden, equilibrated with water. Fractions of 12 ml volume were collected. The wavelength of maximum optical absorption in the ultraviolet region and the optical density at such wavelength were determined for each fraction. Also, the optical density at 340 nm of each fraction after reduction with alcohol dehydrogenase was determined. The fractions which had an optical absorption maximum at 264 nm and had a ratio of optical density at 340 nm to that at 265 nm greater than 0.05 were pooled. The pooled material was concentrated to from 15 to 20 ml on a rotary evaporator and passed through a 2.5×28 cm column of Dowex 1-X8, available from Bio-Rad Laboratories, Richmond, Calif., equilibrated with water. Additional water was added to wash the pooled material through the column, and 10 ml fractions were collected. The fractions which had an optical density at 340 nm to that at 264 nm greater than 0.1 were pooled.

The pooled material was passed through a 5×45 cm column of Dowex 50-X2, available from Bio-Rad laboratories, Richmond, Calif., equilibrated with water. Additional water was added to wash the pooled material through the column and 20 ml fractions were collected. The fractions which had an optical absorption maximum at 264 nm and had a ratio of optical density at 340 nm to that at 264 nm greater than 0.18 were pooled. The pooled material was concentrated to from 4 to 5 ml and purified by electrophoresis as follows.

The concentrated material was applied to a sheet of Whatman 3 MM paper, available from Reeve Angel, Clifton, New Jersey, in a 1 to 2 cm wide strip perpendicular to the direction of current flow. The paper was then wetted with 0.02 M sodium phosphate at pH 6.0. Electrophoresis was conducted according to the Durum hanging paper method, as described in *Science* 121:829(1955), for 4-7 hours with a potential gradient of about 8.5 volts/cm. The location of the desired pyridine nucleotide derivative was determined by fluorescence developed after spraying a test strip of the paper with 0.5 M sodium cyanide according to the procedure described in *J. Biol. Chem.* 191:447(1951). The area containing the desired derivatives was cut out of the paper and extracted with three (3) 50 ml volumes of water. The resulting extracts containing nicotinamide 6-(2-

aminoethylamine) purine dinucleotide were pooled, concentrated to from 3 to 4 ml, and stored at -20° C.

### EXAMPLE 2

#### Preparation of nicotinamide adenine dinucleotide-biotin conjugate

A 16 mg quantity of biotin was suspended in 1 ml of water containing 22 mg of nicotinamide 6-(2-aminoethylamino) purine dinucleotide prepared as in Example 1. A few drops of 0.1 N sodium hydroxide was added to aid dissolution of the biotin. A 240 mg quantity of 1-cyclohexyl-3-(2-morpholinoethyl)-carbodiimide metho-p-tolulene sulfonate was added to the resulting solution and brought into solution by dropwise addition of 0.1 N hydrochloric acid. The reaction mixture was allowed to incubate at room temperature for 5 hours and was then poured into 10 ml of acetone at 31 10° C. The oil which formed was separated, washed twice with from 5 to 10 ml of ether and dissolved in from 1 to 2 ml of water. The resulting material was purified by electrophoresis on paper as in Example 1. Two fluorescent bands appeared after spraying with sodium cyanide, one having migrated toward the cathode and the other toward the anode. The latter band, which contained the NAD-biotin conjugate, was eluted with water and stored at -20° C.

### EXAMPLE 3

#### Preparation of biotin-umbelliferone conjugate

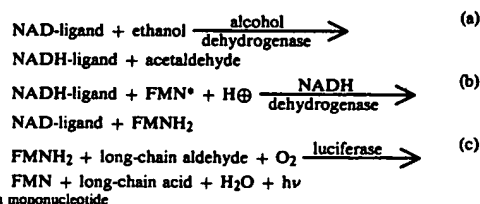
(2-Oxo-2H-1-benzopyran-7-yl)-5-[cis-hexahydro-2-oxo-1H-thieno-(3,4-d)-imidazole]valeric acid ester

A solution of 300 mg (1.2 mmol) anhydrous biotin in 20 ml dry dimethylformamide was stirred at -10° C. under dry nitrogen gas and 0.17 ml (1.2 mmol) dry triethylamine was added. A solution of freshly distilled ethyl chloroformate (0.141 ml in 3 ml of dry ether) was added dropwise. After incubation for 30 min with stirring, the resulting precipitate was filtered under a dry nitrogen atmosphere and cooled immediately to -10° C. To the filtered residue was added a solution of 197 mg (1.2 mmol) anhydrous 7-hydroxycoumarin in 3 ml dry pyridine and stirred for 1 hour at -10° C. followed by 2C hours at 25° C. The solvents were evaporated under high vacuum at 40° C. After cooling, the resulting solid was filtered and recrystallized from methanol to yield the desired product (melting point=216°-218° C.). Calculated for C<sub>19</sub>H<sub>20</sub>N<sub>2</sub>P<sub>5</sub>S: C,48.75; H,4.19; N,7.21. Found: C,58.4; H,5.12; N,6.86.

### EXAMPLE 4

Competitive binding-bioluminescence assay for biotin; effect of varying levels of biotin on the peak light intensity produced.

The bioluminescence reaction system used in this Example was based on the following reactions:



\*flavin mononucleotide

## A. Preparation of light-generating solution

A light-generating solution for carrying out reactions (b) and (c) was prepared as follows. A reagent mixture was prepared containing 0.13 M phosphate buffer at pH 7.0, 0.67 wt% bovine serum albumin, 15.7  $\mu$ M flavin mononucleotide (FMN), and 13.3 mM sodium acetate, and this mixture was stored in the dark at  $-20^{\circ}$  C. An emulsion of 5  $\mu$ l of dodecanal in 5 ml of water was prepared the day the light-generating solution was to be used. Lyophilized luciferase extracted from *Photobacterium fischeri* (enzyme available from Worthington Biochemical Corp., Freehold, N.J.) was added to 0.013 M phosphate buffer at pH 7.3 to a concentration of 20 mg/ml. After 30 minutes the resulting suspension was centrifuged at 1500 xg for 10 minutes and the pellet was discarded. The light-generating solution was then prepared within 5 minutes of use by combining 75  $\mu$ l of the reagent mixture, 5  $\mu$ l of the dodecanal emulsion, and 20  $\mu$ l of the luciferase solution.

## B. Preparation of insolubilized binding partner

Avidin, which has a binding affinity for biotin, was insolubilized by being covalently bound to a water insoluble polymer bead as follows. A quantity of Sepharose 4B (available from Pharmacia AB, Uppsala, Sweden) was activated for bonding to avidin using the method of March et al, *Analytical Biochemistry* 60:149(1974). Approximately 4 ml of the activated Sepharose 4B was suspended in 8 ml of 0.1 M citrate buffer at pH 7.0. To the suspension was added 6 mg of avidin, having an activity of 9.9 units/mg, in 3 ml of water. One unit of avidin activity is that quantity of avidin capable of binding 1  $\mu$ g of biotin. The resulting reaction mixture was stirred for 6 hours at  $7^{\circ}$  C. The avidin-bound-Sepharose 4B was then filtered, washed with 100 ml of 0.1 M sodium bicarbonate buffer at pH 9.0, and resuspended in 240 ml of 0.1 M tris-(hydroxymethyl)-aminomethane hydrochloride buffer at pH 8.0.

## C. Control experiments

Nine specific binding reaction mixtures were prepared, each having a total volume of 0.19 ml and each containing 0.1 M tris-(hydroxymethyl)-aminomethane hydrochloride buffer at pH 8.0, 0.6 M ethanol, 0.01 M semicarbazide hydrochloride, and respectively the amounts or concentrations indicated in Table 1 of NAD, NAD-biotin conjugate, avidin-bound Sepharose 4B suspension (prepared according to Part B of this Example), and Sepharose 4B suspension (formed by suspending 1 ml of packed Sepharose 4B in 60 ml of 0.1 M tris-(hydroxymethyl) aminomethane hydrochloride buffer at pH 8.0). The reaction mixtures were shaken gently for 15 minutes at room temperature. Then, 0.22 International units of alcohol dehydrogenase was added to each reaction mixture to initiate the reduction reaction. Semicarbazide combines with the acetaldehyde produced in reaction (a) to form a semicarbazone and thus to drive reaction (a) in the desired direction.

The reaction mixtures were shaken again for 15 minutes at room temperature. A 10  $\mu$ l aliquot of the supernatant from each reaction mixture was then injected into a separate cuvette mounted in a DuPont Model 760 Bioluminescence Photometer (E. I. duPont de Nemours, Wilmington, Del.) containing 100  $\mu$ l of the previously prepared light-generating solution which had been pre-incubated for from 2 to 3 minutes at  $25^{\circ}$  C. The results appear in Table 1.

TABLE 1

re- ac- tion	concent- ration of NAD(nM)	concent- ration of NAD-bi- otin conju- gate (nM)	avidin- bound Sepharose 4B sus- pension ( $\mu$ l)	Sepharose 4B sus- pension ( $\mu$ l)	peak light inten- sity
1	—	—	—	—	1.2
2	21	—	—	—	159
3	21	—	20	—	147
4	—	10	—	—	45.9
5	—	21	—	—	110
6	—	21	20	—	28.5
7	21	—	—	10	154
8	—	21	—	10	114
9	—	—	20	—	1.6

The results of control reactions 1 and 9 show that in the absence of NAD and NAD-biotin conjugate very little light was produced. Reactions 2 and 3 yielded results indicating that the light producing reaction occurred when free NAD was added and that such reaction was substantially unaffected by the presence of avidin-bound-Sepharose 4B. The results of reactions 4, 5, and 6 show that the NAD-biotin conjugate was active in the light producing reaction, that the peak light intensity produced increased as more NAD-biotin conjugate was present, and that the presence of avidin-bound-Sepharose 4B inhibited light production. Comparison of the results of reactions 3 and 5 with those of 7 and 8 shows that the light producing reaction was not affected by the presence of plain Sepharose 4B.

## D. Assay method

Five additional specific binding reaction mixtures were prepared, each having a volume of 0.19 ml and each containing 0.1 M tris-(hydroxymethyl)-aminomethane hydrochloride buffer at pH 8.0, 0.6 M ethanol, 0.01 M semicarbazide, and respectively the amounts or concentrations indicated in Table 2 of NAD-biotin conjugate, free biotin, and avidin-bound-Sepharose 4B suspension. Each reaction mixture was treated in the same manner as the control reaction mixtures in Part C of this Example. The results appear in Table 2.

TABLE 2

re- ac- tion	concent- ration of NAD-bi- otin conju- gate (nM)	concent- ration of biotin (nM)	avidin- bound Sepharose 4B sus- pension ( $\mu$ l)	peak light inten- sity
10	21	—	—	79.1
11	21	—	20	17.4
12	21	79	20	43.1
13	21	158	20	59.9
14	21	158	—	79.7

The results of reactions 11, 12, and 13 show that free biotin and NAD-biotin conjugate compete effectively for the binding sites on the insolubilized avidin since the peak light intensity produced was dependent upon the amount of free biotin present. Reactions 10 and 14 gave results indicating that in the absence of insolubilized avidin, the peak light intensity produced was constant for vastly different concentrations of free biotin.

It was thus demonstrated in this Example that the amount of NAD-biotin conjugate in the liquid phase was inversely related to the amount of free biotin present and thus the assay method and means of the present

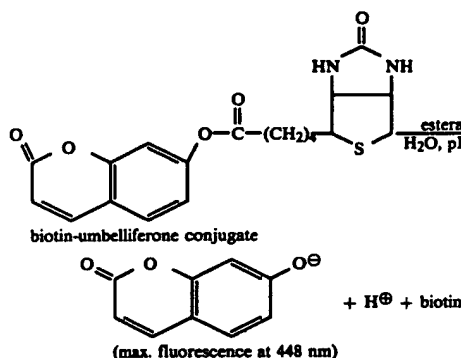


invention are useful in the determination of a ligand in an unknown liquid sample.

### EXAMPLE 3

Specific binding assays for avidin and biotin employing an enzyme substrate as labeling substance.

The specific binding assay systems used in this Example were based on the following reaction:



#### A. Preparation of insolubilized binding partner

Avidin was insolubilized by being covalently bound to a water insoluble polymer bead as in Part B of Example 4 except that after washing with 100 ml of 0.1 M sodium bicarbonate buffer at pH 9.0, the avidin-bound-Sepharose 4B was suspended in 12 ml of 0.1 M tris-(hydroxymethyl)-aminomethane hydrochloride buffer at pH 8.0 and diluted 1:1 with 0.1 M bis-hydroxyethylglycine hydrochloride buffer at pH 7.0.

#### B. Competitive binding assay for biotin; effect of various levels of biotin on the amount of umbelliferone liberated

Eight specific binding reaction mixtures were prepared, each having a total volume of 0.2 ml and each containing 0.1 M bis-hydroxyethylglycine hydrochloride buffer at pH 7.0, 0.3  $\mu$ M biotin-umbelliferone conjugate (prepared as in Example 3), 15  $\mu$ l of the avidin-bound-Sepharose 4B suspension prepared as in Part A of this Example, and biotin in the concentrations indicated in Table 3. The reaction mixtures were allowed to incubate at room temperature with gentle shaking for 20 minutes. Each reaction mixture was centrifuged and a 100  $\mu$ l aliquot of the supernatant was combined with 2 ml of 0.1 M bis-hydroxyethylglycine hydrochloride buffer at pH 8.2 containing 1.08 units of porcine esterase. After a 5 minute incubation at room temperature, the fluorescence intensity produced in each reaction mixture at 448 nm with excitation at 364 nm was measured using an Amico-Bowman spectrophotofluometer. The results appear in Table 3.

TABLE 3

reaction mixture	concentration of biotin ( $\mu$ M)	fluorescence intensity
1	0.00	0.355
2	0.10	0.495
3	0.20	0.469
4	0.30	0.503
5	0.40	0.547
6	0.50	0.502
7	0.75	0.580

TABLE 3-continued

reaction mixture	concentration of biotin ( $\mu$ M)	fluorescence intensity
8	1.00	0.688

It was thus demonstrated in this Example that the amount of NAD-biotin in the liquid phase was directly proportional to the amount of free biotin present and thus the assay method and means of the present method are useful in the determination of a ligand in an unknown liquid sample.

What is claimed is:

1. In a heterogeneous specific binding assay method for determining a ligand in a liquid medium, which method comprises the steps of:

(a) contacting said liquid medium with reagent means including a labeled conjugate comprising a specific binding substance coupled to a labeling substance, said reagent system producing

(1) a bound-phase of the labeled conjugate in which the specific binding substance therein is bound by a specific binding partner thereto and

(2) a free-phase of the labeled conjugate in which the specific binding substance therein is not bound by a specific binding partner thereto,

(b) separating said bound-phase and said free-phase; and

(c) determining said labeling substance in said bound-phase or said free-phase as a function of the amount of said ligand in said liquid medium;

the improvement wherein said labeling substance in said labeled conjugate is a nucleotide coenzyme, and wherein said labeling substance measuring the extent to which said coenzyme participates in an enzymatic reaction involving an enzyme which requires said coenzyme for activity.

2. A method as in claim 1 wherein said nucleotide coenzyme is selected from the group consisting of the adenosine phosphates, nicotinamide adenine dinucleotide and reduced forms thereof, and nicotinamide adenine dinucleotide phosphate and reduced forms thereof.

3. A method as in claim 1 wherein said nucleotide coenzyme is nicotinamide adenine dinucleotide or a reduced form thereof.

4. A method as in claim 1 wherein said nucleotide coenzyme is adenosine triphosphate.

5. A method as in claim 1 wherein said nucleotide coenzyme is flavin adenine dinucleotide.

6. A method as in claim 1 wherein said enzymatic reaction is cyclic.

7. A method as in claim 6 wherein said coenzyme-active labeling substance is a cycled reagent in said cyclic reaction.

8. A method as in claim 1 wherein said enzymatic reaction is chemiluminescent.

9. A method as in claim 1 wherein said ligand is selected from the group consisting of antigens and antibodies thereto; haptens and antibodies thereto; and hormones, vitamins, metabolites, and pharmacological agents, and their receptors and binding substances.

10. A method as in claim 1 wherein said liquid medium is a biological fluid.

11. In a reagent means for use in determining a ligand in a liquid medium, which means comprises (i) a labeled conjugate comprising said ligand or a specific binding analog thereof coupled to a labeling substance, and (ii) a specific binding partner of said ligand which binding

partner is in a form which is insoluble in said liquid medium,

the improvement wherein said labeling substance in said labeled conjugate is a nucleotide coenzyme.

12. Means as in claim 11 wherein said nucleotide coenzyme is selected from the group consisting of the adenosine phosphates, nicotinamide adenine dinucleotide and reduced forms thereof, and nicotinamide adenine dinucleotide phosphate and reduced forms thereof.

13. Means as in claim 11 wherein said nucleotide coenzyme is nicotinamide adenine dinucleotide or a reduced form thereof.

14. Means as in claim 11 wherein said nucleotide coenzyme is adenosine triphosphate.

15. Means as in claim 11 wherein said nucleotide coenzyme is flavin adenine dinucleotide.

16. Means as in claim 11 wherein said ligand is selected from the group consisting of antigens and antibodies thereto; haptens and antibodies thereto; and hormones, vitamins, metabolites, and pharmacological agents, and their receptors and binding substances.

17. Means as in claim 11 wherein at least one of the components of said means is incorporated with a carrier that is insoluble in said liquid medium.

18. Means as in claim 17 wherein said carrier is a matrix that is absorbent relative to said liquid medium.

19. In a reagent means for use in determining a ligand in a liquid medium, which means comprises (i) a labeled conjugate comprising a specific binding partner of said

ligand coupled to a labeling substance, and (ii) a specific binding partner of said ligand which binding partner is in a form which is insoluble in said liquid medium,

the improvement wherein said labeling substance in said labeled conjugate is a nucleotide coenzyme.

20. Means as in claim 19 wherein said nucleotide coenzyme is selected from the group consisting of the adenosine phosphates, nicotinamide adenine dinucleotide and reduced forms thereof, and nicotinamide adenine dinucleotide phosphates and reduced forms thereof.

21. Means as in claim 19 wherein said nucleotide coenzyme is nicotinamide adenine dinucleotide or a reduced form thereof.

22. Means as in claim 19 wherein said nucleotide coenzyme is adenosine triphosphate.

23. Means as in claim 19 wherein said nucleotide coenzyme is flavin adenine dinucleotide.

24. Means as in claim 19 wherein said ligand is selected from the group consisting of antigens and antibodies thereto; haptens and antibodies thereto; and hormones, vitamins, metabolites, and pharmacological agents, and their receptors and binding substances.

25. Means as in claim 19 wherein at least one of the components of said means is incorporated with a carrier that is insoluble in said liquid medium.

26. Means as in claim 25 wherein said carrier is a matrix that is absorbent relative to said liquid medium.

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UNITED STATES PATENT AND TRADEMARK OFFICE  
**CERTIFICATE OF CORRECTION**

PATENT NO. : 4,230,797

DATED : October 28, 1980

INVENTOR(S) : R. C. Boguslaski et al

It is certified that error appears in the above-identified patent and that said Letters Patent are hereby corrected as shown below:

Col. 13, line 42, delete "2" appearing before "ATP"

Claim 1, line 21, after "labeling substance"  
insert --is determined in  
said bound-phase or said  
free-phase by--

Claim 20, line 5, "phosphates" should read "phosphate"

**Signed and Sealed this**

*Tenth Day of February 1981*

[SEAL]

*Attest:*

RENE D. TEGTMEYER

*Attesting Officer*

*Acting Commissioner of Patents and Trademarks*

